

## Protective effects of benidipine on hydrogen peroxide-induced injury in rat isolated hearts

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### Abstract

We investigated the effects of benidipine (hydrochloride), a calcium antagonist, on hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>)-induced injury in Langendorff-perfused rat hearts. The hearts were aerobically perfused at a constant flow and exposed to H<sub>2</sub>O<sub>2</sub> (600 μmol L<sup>-1</sup>) for 4 min, resulting in the oxidative stress-induced myocardial dysfunction (e.g., decrease in the left ventricular developed pressure) and myocardial cell injury (e.g., increase in the release of lactate dehydrogenase). Pretreatment of the hearts with benidipine or nifedipine was performed for 20 min until the start of H<sub>2</sub>O<sub>2</sub> exposure. Benidipine at 1 nmol L<sup>-1</sup> and nifedipine at 10 nmol L<sup>-1</sup> decreased the myocardial contractility and perfusion pressure to a similar degree in the hearts under normal conditions. Benidipine (1 nmol L<sup>-1</sup>) significantly reduced the H<sub>2</sub>O<sub>2</sub>-induced myocardial damage. Nifedipine (10 nmol L<sup>-1</sup>) also tended to exhibit similar effects. Benidipine inhibited the increase in tissue lipid peroxidation induced by H<sub>2</sub>O<sub>2</sub>. The results suggest that, in addition to the calcium antagonism, benidipine possesses other actions responsible for the cardioprotective effects, to which the antioxidant activity of benidipine may partly contribute.

### Introduction

Benidipine (hydrochloride) is a 1,4-dihydropyridine calcium antagonist with a slow onset of action and long-lasting properties (Karasawa & Kubo 1988a; Karasawa et al 1988a), which is clinically useful in the treatment of hypertension and angina pectoris (Fuji et al 1988; Yamada et al 1990). In experimental models, benidipine exhibited beneficial effects against ischaemic and reperfusion injury of the heart (Karasawa & Kubo 1988b; Karasawa et al 1988b). Myocardial ischaemia and reperfusion induces intracellular calcium overload, resulting in myocardial injury (Kloner & Braunwald 1987). The inhibition of calcium currents is probably the main mechanism for the cardioprotective action of benidipine.

During reperfusion after ischaemia, reactive oxygen species (known as potent oxidants), such as superoxide anion, hydroxyl radical and hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), are generated and cause lipid peroxidation of the cell membrane and intracellular Ca<sup>2+</sup> accumulation, which induces mechanical and metabolic derangements or damage (Kaneko et al 1994). Antioxidants, as well as oxygen radical scavengers, protect the myocardium against ischaemia–reperfusion damage (Hess & Manson 1984; Lucchesi 1990; Loesser et al 1991). Dihydropyridine calcium antagonists, including benidipine, have an antioxidant property (Mason & Trumbore 1996; Sugawara et al 1996; Yao et al 2000). In fact, benidipine has been shown to preserve the vascular endothelial function in rat arteries subjected to ischaemia and reperfusion (Karasawa et al 1991; Yao & Karasawa 1994), which was independent of the calcium-channel blocking action (Sato et al 1992). Thus, the antioxidant action is expected to contribute to the cardioprotective effect of benidipine. We reported that benidipine prevented myocardial injury following ischaemia and reperfusion in rat isolated perfused hearts (Yao & Karasawa 1994). In this study, to evaluate the contribution of the antioxidant action of

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benidipine to its cardioprotection in the therapeutic range of concentrations, we examined the effects of benidipine on the H<sub>2</sub>O<sub>2</sub>-induced myocardial injury in rat isolated hearts.

## Materials and Methods

### Drugs and chemicals

Benidipine hydrochloride was synthesized at Kyowa Hakko Kogyo. Nifedipine was purchased from the Sigma Chemical Co. (St Louis, MO). Benidipine and nifedipine were initially dissolved in polyethylene glycol 400 (Kanto Chemical, Tokyo, Japan) to make a 1 mmol L<sup>-1</sup> solution, and the solution was further diluted with distilled water before use in the experiments. All other chemicals were of reagent grade and were from commercial sources.

### Animals

Male Wistar rats, 300–390 g (Japan SLC Inc., Hamamatsu, Japan), were used. The rats were kept at 19–25°C under a 12-h light–dark cycle, and they had free access to tap water and commercial chow (FR-2, Funabashi Farm, Funabashi, Japan). All rats received humane care in compliance with the Guiding Principles for the Care and Use of Laboratory Animals, formulated by the Japanese Pharmacological Society, and the protocol was approved by the Bioethical Committee of Pharmaceutical Research Institute, Kyowa Hakko Kogyo Co. Ltd.

### Heart perfusion

Rats were anaesthetized with sodium pentobarbital (Nembutal 50 mg kg<sup>-1</sup>, i.p.; Dainippon Pharmaceutical, Osaka, Japan) 30 min after an injection with heparin (500 IU/rat, i.p.). Under anaesthesia, the chest was opened and the heart was excised. The aorta was cannulated, and the heart was perfused at a constant flow of 10 mL min<sup>-1</sup> with oxygenated (95% O<sub>2</sub>+5% CO<sub>2</sub>) Krebs–Henseleit bicarbonate (KHB) buffer (25°C) with the following composition (mmol L<sup>-1</sup>): NaCl, 112; NaHCO<sub>3</sub>, 25; KCl, 5; MgSO<sub>4</sub>, 1.2; KH<sub>2</sub>PO<sub>4</sub>, 1; CaCl<sub>2</sub>, 1.25; dextrose, 11.5; pyruvate, 2. A balloon-tipped catheter filled with water was then inserted into the left ventricle via the left atrium. The heart was mounted on a Langendorff apparatus and perfused with warmed KHB buffer (37°C). Perfusion pressure (PP) and left ventricular pressure (LVP) were measured with the aortic cannula and the balloon-tipped catheter, which were connected to pressure transducers (SCK-590, Nihon Kohden, Tokyo, Japan), respectively. The maximum rate of rise in LVP (LV dp/dt max) was derived using an electric differentiator (ED-600G, Nihon Kohden). All parameters were continuously recorded on a thermal array chart recorder (WS-681, Nihon Kohden). The left ventricular end-diastolic pressure (LVEDP) was measured from the recorded LVP signal.

The heart was allowed to equilibrate for 15 min, and then LVEDP was adjusted to about 5 mmHg by changing the volume of the balloon. The left ventricular developed

pressure (LVDP) was calculated from the difference between left ventricular peak systolic pressure and LVEDP.

### Experimental protocol

Once the baseline values were measured, the hearts were treated with benidipine (hydrochloride, 0.1–1 nmol L<sup>-1</sup>), nifedipine (1–10 nmol L<sup>-1</sup>) or vehicle (0.001% polyethylene glycol 400) 20 min before H<sub>2</sub>O<sub>2</sub> treatment. The concentrations of benidipine were within its therapeutic range. The concentrations of nifedipine were selected to cause a similar degree of negative inotropic effects under normal conditions. The drug solution or the vehicle was infused into the perfusion line leading to the aortic cannula using a constant flow infusion pump (Pump 22; Harvard Apparatus Inc., South Natick, MA) at a rate of 0.1 mL min<sup>-1</sup> for 20 min. After the post-drug cardiac functions were measured and the perfusate was collected for 1 min, H<sub>2</sub>O<sub>2</sub> solution (Kokusan Chemical Works, Tokyo, Japan) was infused into the aortic cannula at a rate of 0.1 mL min<sup>-1</sup> for 4 min. The final concentration of H<sub>2</sub>O<sub>2</sub> in the perfusate was 600 μmol L<sup>-1</sup>. The heart was then perfused with control buffer for 30 min. At 0, 10, 20 and 30 min after stoppage of H<sub>2</sub>O<sub>2</sub> infusion, the perfusate was sampled for 1 min. The activity of lactate dehydrogenase (LDH) in the perfusate samples was measured with an autoanalyser (AU600; Olympus, Tokyo, Japan).

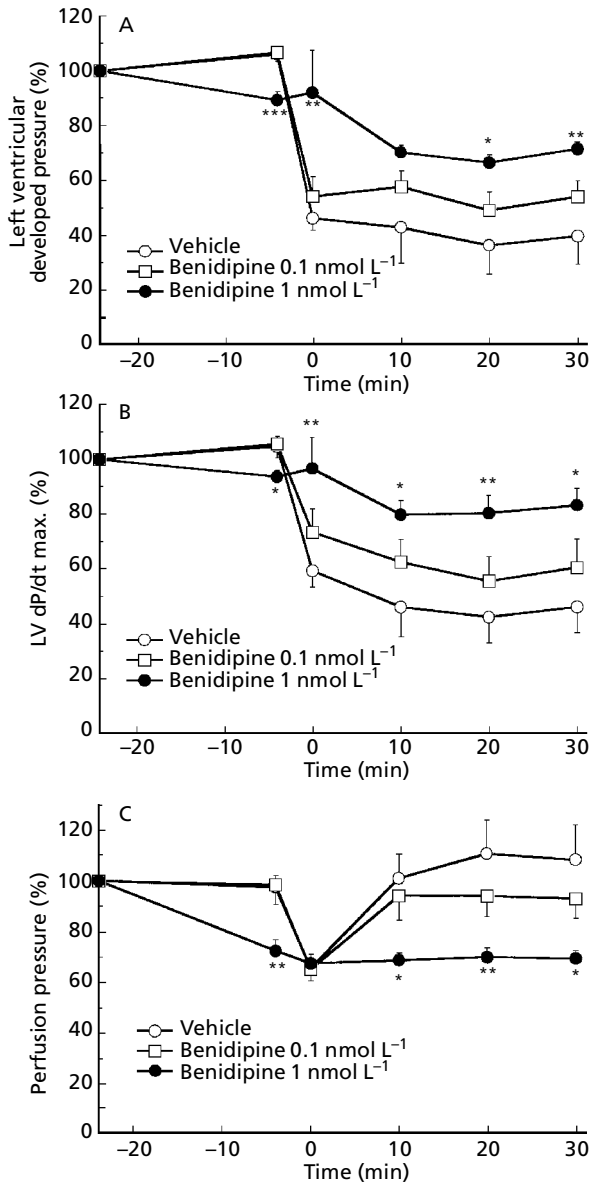
### Measurement of lipid peroxidation

To measure the tissue levels of lipid peroxidation in normal, H<sub>2</sub>O<sub>2</sub>-treated and benidipine+H<sub>2</sub>O<sub>2</sub>-treated hearts, the hearts were frozen at the end of the experiment with freezing clamps previously chilled in liquid nitrogen. The frozen myocardial samples were stored at –80°C until the analysis was performed.

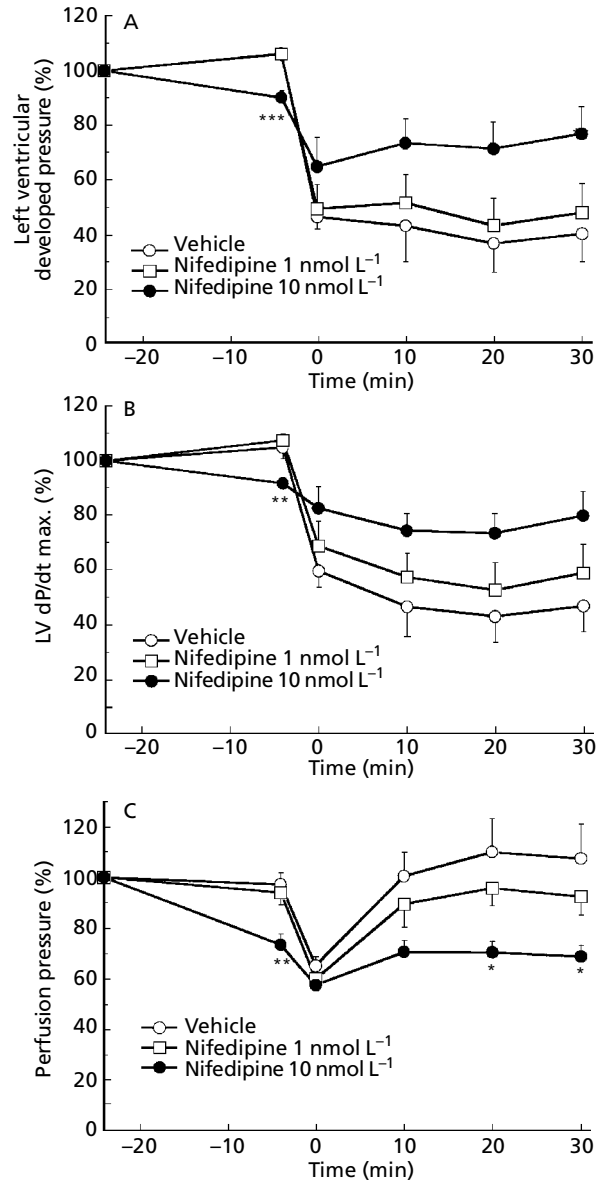
The samples were pulverized in a mortar cooled with liquid nitrogen. A part of the pulverized tissue powder (about 100 mg) was weighed and homogenized 1:10 (w/v) in 10% KCl. Levels of thiobarbituric acid reactive substances (TBARS) were determined by the method of Ohkawa et al (1979) as an index of the extent of lipid peroxidation. TBARS formed in the solution were determined after heating at 100°C for 60 min in the presence of 0.2% thiobarbituric acid in a total volume of 4.0 mL. The TBARS were extracted into n-butanol and pyridine (15:1, v/v). The fluorescence was measured at an excitation wavelength of 515 nm and an emission wavelength of 553 nm (F-3000, Hitachi, Tokyo).

### Statistical analysis

All values are expressed as means ± s.e.m. The drug-treatment values of the parameters measured were compared with the vehicle-treatment values. Inter-group comparisons were made by the nested variance analysis, and then significant differences were tested by one-way analysis of variance followed by Dunnett's test or the Kruskal–Wallis test followed by Steel's test for multiple comparison, or the



**Figure 1** Effects of benidipine on left ventricular developed pressure (LVDP, Panel A), maximal rate of rise in left ventricular pressure (LV dP/dt max, Panel B) and perfusion pressure (PP, Panel C) in rat isolated perfused hearts subjected to  $H_2O_2$ . The hearts were treated with benidipine (0.1 or 1  $nmol L^{-1}$ ) for 20 min until the start of  $H_2O_2$  infusion. The ordinate indicates percentage of the pre-drug value. Values are means  $\pm$  s.e.m. of 6 or 7 hearts. \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$  vs vehicle. Basal values of LVDP, LV dP/dt max and PP in each group were as follows. LVDP (mmHg): control,  $107.2 \pm 3.4$ ; benidipine 0.1  $nmol L^{-1}$ ,  $104.0 \pm 6.5$ ; and benidipine 1  $nmol L^{-1}$ ,  $99.1 \pm 3.4$ . LV dP/dt max ( $mmHg s^{-1}$ ): control,  $3329 \pm 174$ ; benidipine 0.1  $nmol L^{-1}$ ,  $2983 \pm 193$ ; and benidipine 1  $nmol L^{-1}$ ,  $2980 \pm 144$ . PP (mmHg): control,  $68.4 \pm 6.9$ ; benidipine 0.1  $nmol L^{-1}$ ,  $61.9 \pm 5.6$ ; and benidipine 1  $nmol L^{-1}$ ,  $67.3 \pm 6.5$ . No significant difference was observed among all the parameters in each group.



**Figure 2** Effects of nifedipine on left ventricular developed pressure (LVDP, Panel A), maximal rate of rise of left ventricular pressure (LV dP/dt max, Panel B) and perfusion pressure (PP, Panel C) in rat isolated perfused hearts subjected to  $H_2O_2$ . The hearts were treated with nifedipine (1 or 10  $nmol L^{-1}$ ) for 20 min until the start of  $H_2O_2$  infusion. The ordinate indicates the percentage of the pre-drug value. Values are means  $\pm$  s.e.m. of 6 or 7 hearts. \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$  vs vehicle. Basal values of LVDP, LV dP/dt max and PP in each group were as follows. LVDP (mmHg): control,  $107.2 \pm 3.4$ ; nifedipine 1  $nmol L^{-1}$ ,  $97.8 \pm 5.1$ ; and nifedipine 10  $nmol L^{-1}$ ,  $95.2 \pm 3.2$ . LV dP/dt max ( $mmHg s^{-1}$ ): control,  $3329 \pm 174$ ; nifedipine 1  $nmol L^{-1}$ ,  $2991 \pm 100$ ; and nifedipine 10  $nmol L^{-1}$ ,  $2996 \pm 209$ . PP (mmHg): control,  $68.4 \pm 6.9$ ; nifedipine 1  $nmol L^{-1}$ ,  $67.9 \pm 6.8$ ; and nifedipine 10  $nmol L^{-1}$ ,  $71.1 \pm 5.2$ . No significant difference was observed among all the parameters in each group.

F test followed by Student's *t*-test for non-matched pair comparison. A difference was considered as statistically significant at  $P < 0.05$ . All statistics were calculated with

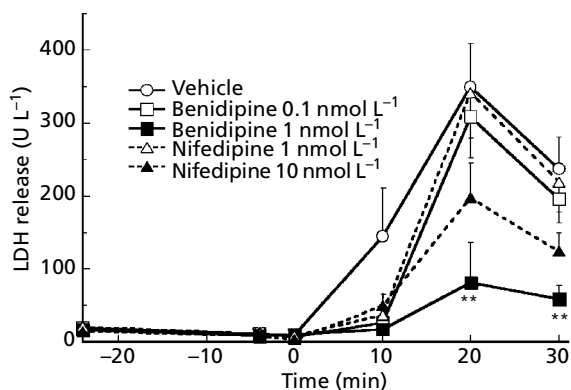
the SAS statistical software (version 6.12; SAS Institute Inc., Cary, NC).

## Results

Figures 1 and 2, respectively, show the effects of benidipine and nifedipine on the  $H_2O_2$ -induced changes in LVDP, LV dP/dt max and PP. There were no significant differences in the baseline values among the groups. Benidipine at  $1 \text{ nmol L}^{-1}$  and nifedipine at  $10 \text{ nmol L}^{-1}$  decreased the LV dP/dt max as an index of myocardial contractility. No significant differences in the effects on any cardiac parameters were detected between benidipine at  $1 \text{ nmol L}^{-1}$  and nifedipine at  $10 \text{ nmol L}^{-1}$  20 min after the beginning of infusion of each drug.  $H_2O_2$  produced contractile failure as demonstrated by the decreases in LVDP and LV dP/dt max.  $H_2O_2$  produced the decrease in PP, indicating vasodilatation in the hearts perfused at a constant flow, at 0 min after the stoppage of  $H_2O_2$  perfusion, and then gradual increase in PP. At 30 min after  $H_2O_2$  perfusion, the mean LVDP value of the hearts in the vehicle group was 45% of the pre- $H_2O_2$  value. Treatment with benidipine at  $1 \text{ nmol L}^{-1}$  significantly inhibited the decreases in LVDP and LV dP/dt max, and the increase in PP. Nifedipine at  $10 \text{ nmol L}^{-1}$  reduced the  $H_2O_2$ -induced changes in LVDP, LV dP/dt max and PP, but the effects were not significant. Benidipine at  $0.1 \text{ nmol L}^{-1}$  and nifedipine at  $1 \text{ nmol L}^{-1}$  did not affect these parameters.

Figure 3 shows the time course of changes in the LDH activity. Perfusion with vehicle alone caused an increase in the LDH release that peaked 20 min after the start of the treatment with  $H_2O_2$ . Benidipine and nifedipine reduced the release of LDH. At  $1 \text{ nmol L}^{-1}$ , the effect of benidipine on the release of LDH was significant.

Hearts treated with  $H_2O_2$  had significantly elevated levels of TBARS ( $16.3 \text{ nmol mL}^{-1}$  10% homogenates,  $P < 0.001$ ) compared with normal hearts ( $10.4 \text{ nmol mL}^{-1}$ ). Benidipine at  $1 \text{ nmol L}^{-1}$  significantly reduced the rise in TBARS ( $12.6 \pm 1.3 \text{ nmol mL}^{-1}$ ,  $P < 0.05$ ).



**Figure 3** Effects of benidipine and nifedipine on the release of lactate dehydrogenase (LDH) in rat isolated perfused hearts subjected to  $H_2O_2$ . The hearts were treated with benidipine ( $0.1$  or  $1 \text{ nmol L}^{-1}$ ) or nifedipine ( $1$  or  $10 \text{ nmol L}^{-1}$ ) for 20 min until the start of  $H_2O_2$  infusion. Values are means  $\pm$  s.e.m. of 6 or 7 hearts. \*\* $P < 0.01$  vs vehicle.

## Discussion

In this study,  $H_2O_2$  produced myocardial membrane damage and subsequent mechanical dysfunction in isolated perfused hearts from rats. Several studies indicated that  $H_2O_2$  decreased the tissue levels of high-energy phosphates in the heart, leading to cardiac dysfunction (Hara et al 1993; Hara & Abiko 1996; Kokita & Hara 1996; Xiao et al 1999). According to recent physiological studies,  $H_2O_2$  activated the  $Na^+/Ca^{2+}$  exchanger through increases in intracellular  $Na^+$  in ventricular myocytes (Yanagida et al 1995; Goldhaber 1996; Ward & Giles 1997). In addition,  $H_2O_2$  was reported to induce the enhancement of the L-type calcium currents in the myocytes (Thomas et al 1998). These effects probably cause the calcium overload, which is responsible for irreversible myocardial derangements in the cardiac myocytes (Goldhaber 1996).

We examined the effects of benidipine and nifedipine on the  $H_2O_2$ -induced myocardial injury in rat isolated perfused hearts. These calcium antagonists protected the hearts against the  $H_2O_2$ -induced injury. In particular, benidipine exhibited a protective effect even at a concentration of  $1 \text{ nmol L}^{-1}$ , which corresponds to the therapeutic plasma concentrations. One  $\text{nmol L}^{-1}$  of benidipine decreased LVDP, LV dP/dt max, and PP. L-type  $Ca^{2+}$ -channel-blocking action, leading to increasing coronary flow and decreasing cardiac mechanical function, may partly contribute to the cardioprotective effects of benidipine. One  $\text{nmol L}^{-1}$  of benidipine, but not  $10 \text{ nmol L}^{-1}$  of nifedipine, significantly prevented the  $H_2O_2$ -induced myocardial injury, even though both decreased the myocardial contractility and perfusion pressure to a similar degree in the hearts under normal conditions. The protective effects of benidipine could not be totally attributed to its inhibition of L-type calcium current. Just after the treatment with  $H_2O_2$  (0 min), LVDP and the LV dP/dt max were decreased without increases in LDH release, indicating that the contractile dysfunction was not secondary to the myocardial cell injury. Benidipine at  $1 \text{ nmol L}^{-1}$  inhibited these decreases at 0 min, suggesting that benidipine may directly prevent the  $H_2O_2$ -induced myocardial dysfunction.

Benidipine has been shown to increase cardiac NO levels and exhibit protective effects against ischaemic damage in canine hearts via bradykinin- and NO-dependent mechanisms (Kitakaze et al 1999b; Asanuma et al 2001). NO donors had beneficial effects on the LVDP in the ischaemia-reperfusion injury of Langendorff hearts (Hotta et al 1999). NO had inhibitory effects on left ventricular dysfunction after  $H_2O_2$ -mediated oxidative stress in rat isolated hearts (Valen et al 1996). NO may contribute to the cardioprotective effects of benidipine in our study.

Ischaemia-reperfusion-induced increases in the level of free radicals are paralleled by elevated rates of lipid peroxidation (Ambrosio et al 1991; Blasig et al 1995; Cordis et al 1995). Some radical scavengers and antioxidants have been demonstrated to protect the myocardium against oxidative stress (Nakaya et al 1987). Several studies indicate that some calcium antagonists possess lipid antioxidant properties (Mason & Trumbore 1996; Sugawara et al 1996; Yao et al 2000). We have also reported that eight dihydro-

pyridine (DHP) calcium antagonists, including benidipine and nifedipine, had concentration-dependent antioxidant effects in a rat brain homogenate (Yao et al 2000). A hydroxyl radical that is generated from  $H_2O_2$  by the Fenton reaction easily removes a hydrogen atom from a methylene carbon of an unsaturated fatty acid of the membrane phospholipids, and hence produces peroxidation of lipids (Hess & Manson 1984). In fact,  $H_2O_2$  increased the tissue level of TBARS, products of lipid peroxidation. It has been demonstrated that myocardial changes induced by reactive oxygen are accompanied by lipid peroxidation of the cell membrane of the heart (Hess & Manson 1984). Antioxidant substances have been shown to attenuate both myocardial dysfunction and lipid peroxidation induced by  $H_2O_2$  (Onodera et al 1992; Nagy et al 1996). In this study, benidipine inhibited the  $H_2O_2$ -induced lipid peroxidation, suggesting that the protective effects of benidipine on the  $H_2O_2$ -induced injury may be due, in part, to its antioxidant effect. However, the exact mechanism remains unknown, and further investigations are necessary.

In general, DHP calcium antagonists appear to be distributed in cell membranes according to their lipophilicity (Mason & Trumbore 1996). Nosaka & Ishii (1991) reported that, in the octanol–buffer system and lipid membranes, benidipine had the highest partition coefficient of the DHP calcium antagonists, including nifedipine. In addition, the membrane approach theory is considered as a putative mechanism of long-lasting antihypertensive effects of benidipine, and explains the lack of correlation between the effects and plasma levels of benidipine (Kitakaze et al 1999a). These results suggest that benidipine is highly lipophilic and effectively accumulates in membranes. In fact, the vasorelaxant effect of benidipine in isolated coronary arteries was sustained, even after washout of the drug (Karasawa & Kubo 1988a). In this study, benidipine may effectively and continuously accumulate in the myocardial membranes, contributing to the cardioprotective efficacy of benidipine.

## Conclusions

Benidipine has protective effects on the  $H_2O_2$ -induced injury in rat isolated hearts. These results suggest that, in addition to calcium antagonism, benidipine possesses other actions responsible for the cardioprotective effects, to which the antioxidant activity of benidipine may partly contribute.

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